

# Neonatal EEG service evaluation

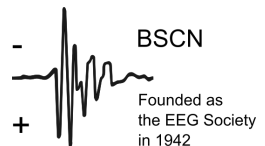
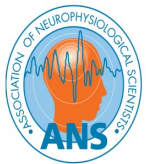
## - Recommendations



### **BSCN and ANS Audit meeting**

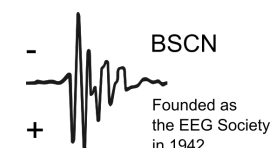
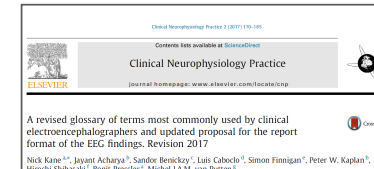
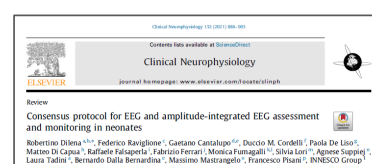
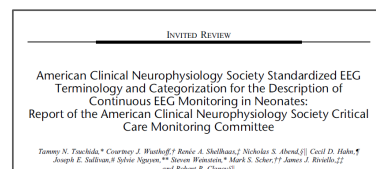
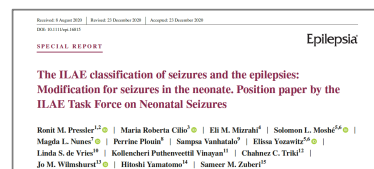
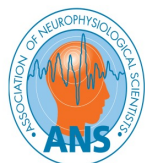
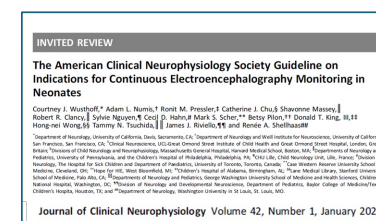
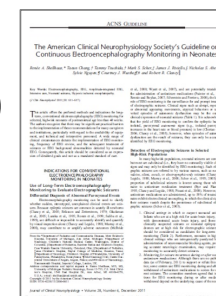
Ronit Pressler, Bryony Carr, Daniela P. Quayle,  
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Birmingham, 2<sup>nd</sup> May 2025



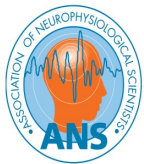
# Recommendations for neonatal EEG monitoring

- ACNS EEG monitoring recommendations (Wusthoff et al 2025)
- Italian Consensus protocol for EEG/aEEG (Dilena et al 2021)
- French technical recommendations (Malfilâtre et al 2021)
- Terminology and reporting of neonatal EEG
  - Tsuchida et al 2013: ACNS glossary
  - Kane et al 2017: IFCN glossary of terminology, 2017 revision
  - Bourel-Ponchel et al 2021: updated French glossary
- Classification of seizures (Pressler et al 2021)



# ACNS GUIDELINE: Indications for cEEG

- Update of the 2011 guidelines (Shellhaas et al 2011)
- Members of ACNS plus patient representative
- Based on Systematic review
  - Seven priority questions
  - Identification of evidence
  - Evaluation of evidence (GRADE)
  - Evaluation on quality of evidence
  - Recommendations



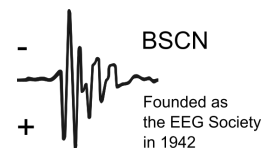
## INVITED REVIEW

### The American Clinical Neurophysiology Society Guideline on Indications for Continuous Electroencephalography Monitoring in Neonates

Courtney J. Wusthoff,\* Adam L. Numis,† Ronit M. Pressler,‡ Catherine J. Chu,§ Shavonne Massey,|| Robert R. Clancy,|| Sylvie Nguyen,¶ Cecil D. Hahn,# Mark S. Scher,\*\* Betsy Pilon,†† Donald T. King, III,‡‡ Hong-wei Wong,§§ Tammy N. Tsuchida,|| James J. Riviello,¶¶ and Renée A. Shellhaas##

\*Department of Neurology, University of California, Davis, Sacramento, CA; †Department of Neurology and Weill Institute for Neuroscience, University of California San Francisco, San Francisco, CA; ‡Clinical Neuroscience, UCL-Great Ormond Street Institute of Child Health and Great Ormond Street Hospital, London, Great Britain; §Divisions of Child Neurology and Neurophysiology, Massachusetts General Hospital, Harvard Medical School, Boston, MA; ¶Departments of Neurology and Pediatrics, University of Pennsylvania, and the Children's Hospital of Philadelphia, Philadelphia, PA; §CHU Lille, Child Neurology Unit, Lille, France; \*\*Division of Neurology, The Hospital for Sick Children and Department of Paediatrics, University of Toronto, Toronto, Canada; ††Case Western Reserve University School of Medicine, Cleveland, OH; ††Hope for HIE, West Bloomfield, MI; ††Children's Hospital of Alabama, Birmingham, AL; ††Lane Medical Library, Stanford University School of Medicine, Palo Alto, CA; ††Departments of Neurology and Pediatrics, George Washington University School of Medicine and Health Sciences, Children's National Hospital, Washington, DC; ††Division of Neurology and Developmental Neuroscience, Department of Pediatrics, Baylor College of Medicine/Texas Children's Hospital, Houston, TX; and ††Department of Neurology, Washington University in St Louis, St. Louis, MO.

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## PICO

1: In neonates presenting with clinically suspected seizures, does cEEG monitoring improve accuracy of diagnosis?

2: In neonates presenting with clinically suspected seizures, does cEEG monitoring improve accuracy of diagnosis as compared to spot EEG alone?

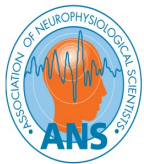
3: In neonates presenting with aEEG events suspicious for seizures, does cEEG monitoring improve accuracy of diagnosis?

4: What is the yield of cEEG monitoring for neonates at risk for seizures in the absence of clinically evident seizures?

5: What is the yield of cEEG monitoring in neonates with definite seizures (whether clinically or by EEG) to assess seizure control after treatment?

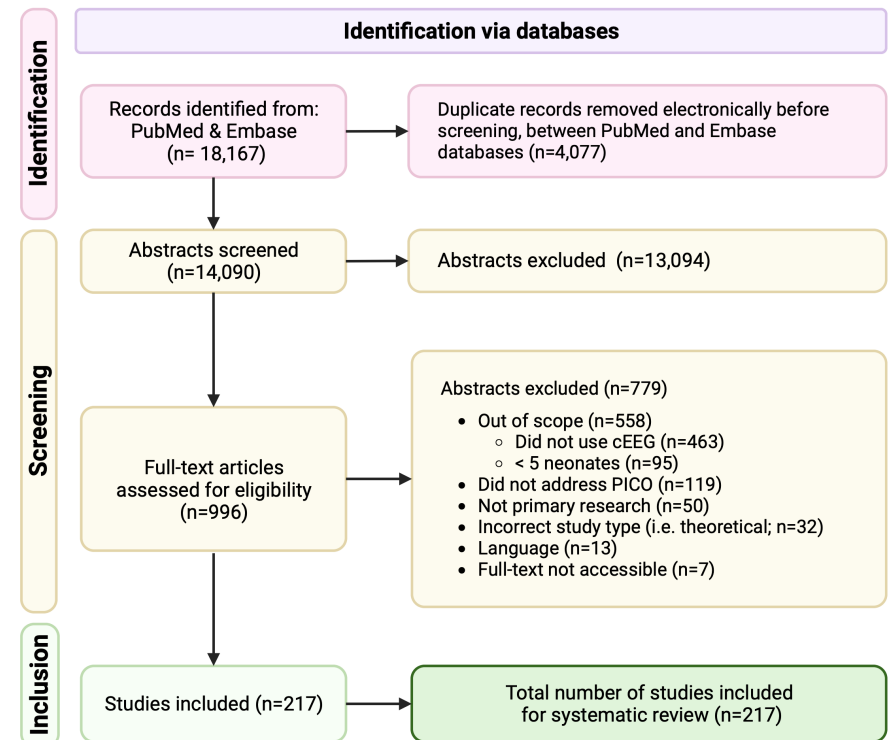
6: What clinically relevant information can be gained from cEEG used as part of the evaluation of encephalopathy?

7: What clinically relevant information can be gained from cEEG used to evaluate brain function in preterm neonates other than for seizures?



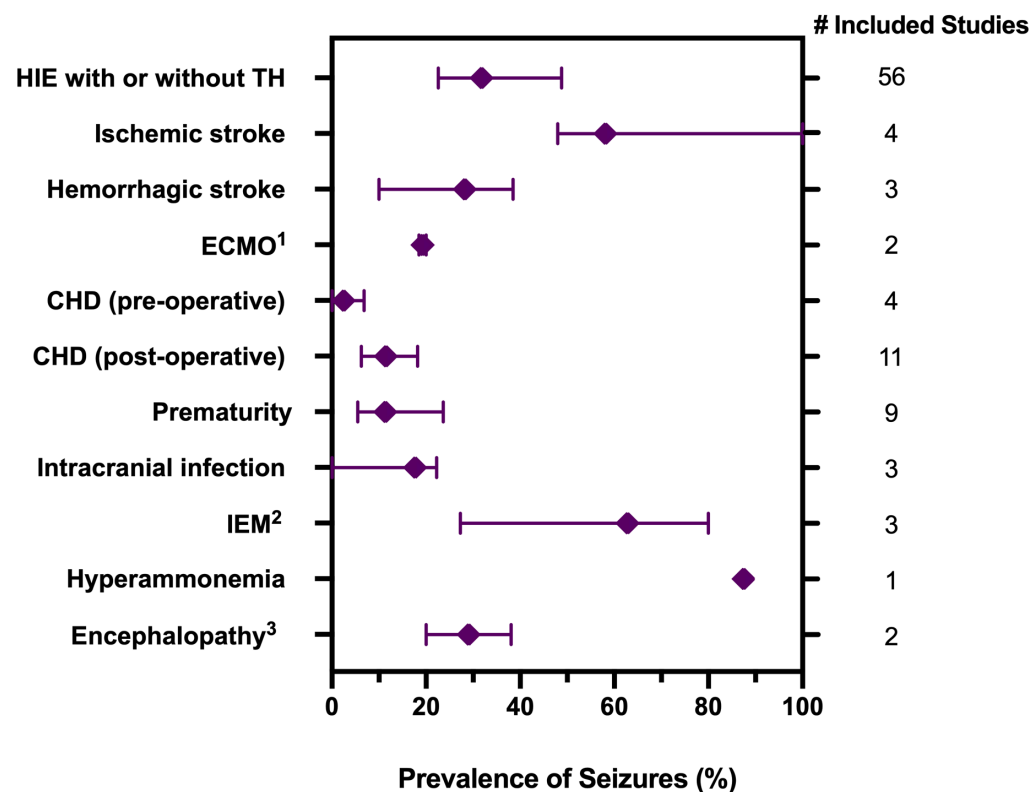
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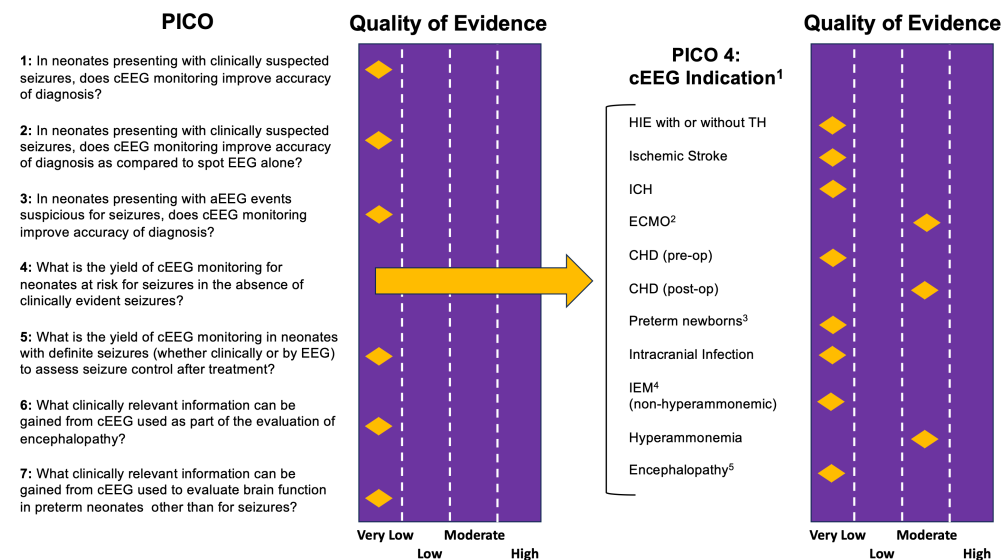
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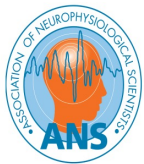
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  - **Evaluation on quality of evidence**
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\*Department of Neurology, University of California, Davis, Sacramento, CA; †Department of Neurology and Weill Institute for Neuroscience, University of California San Francisco, San Francisco, CA; ‡Clinical Neuroscience, UCL-Great Ormond Street Institute of Child Health and Great Ormond Street Hospital, London, Great Britain; §Divisions of Child Neurology and Neurophysiology, Massachusetts General Hospital, Harvard Medical School, Boston, MA; ¶Departments of Neurology and Pediatrics, University of Pennsylvania, and the Children's Hospital of Philadelphia, Philadelphia, PA; §CHU Lille, Child Neurology Unit, Lille, France; \*\*Division of Neurology, The Hospital for Sick Children and Department of Paediatrics, University of Toronto, Toronto, Canada; ††Case Western Reserve University School of Medicine, Cleveland, OH; ††Hope for HIE, West Bloomfield, MI; ††Children's Hospital of Alabama, Birmingham, AL; ††Lane Medical Library, Stanford University School of Medicine, Palo Alto, CA; ††Departments of Neurology and Pediatrics, George Washington University School of Medicine and Health Sciences, Children's National Hospital, Washington, DC; ††Division of Neurology and Developmental Neuroscience, Department of Pediatrics, Baylor College of Medicine/Texas Children's Hospital, Houston, TX; and ††Department of Neurology, Washington University in St Louis, St. Louis, MO.

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# ACNS GUIDELINE: Indications for cEEG

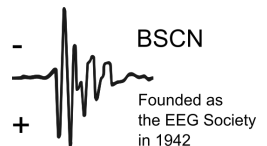
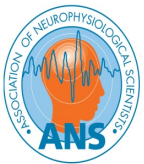
## We suggest cEEG be used

- To improve accuracy of seizure diagnosis in neonates with clinically suspected seizures, as compared to observation alone, aEEG alone, or routine EEG. (Conditional)
- To confirm diagnosis of aEEG events suspected to be seizures. (Conditional)
- For monitoring neonates at risk for seizures without clinically events. (Conditional)
- In neonates with definite seizures to assess for seizure control after treatment and to confirm resolution of seizures. (Conditional)
- There was insufficient evidence to make a recommendation for or against cEEG monitoring after weaning or discontinuing anti-seizure medications.
- among neonates with encephalopathy and preterm neonates for assessment of interictal background to prediction risk of seizures, death or outcome (Conditional)



# BSCN / ANS Neonatal EEG Standards & Guidelines

- Consensus based Standards and Guidelines based on
  - Findings of Neonatal audit
  - Current literature (Tsuchida et al 2013, Kane et al 2017, Bourel-Ponchel et al 2021, Pressler et al 2021, Dilella et al 2021, Mafilâtre et al 2021, Wusthoff et al 2025)
- Consists of
  - INDICATIONS – Ronit Pressler
  - RECORDING STANDARDS – Bryony Carr
  - REPORTING STANDARDS – Ronit Pressler

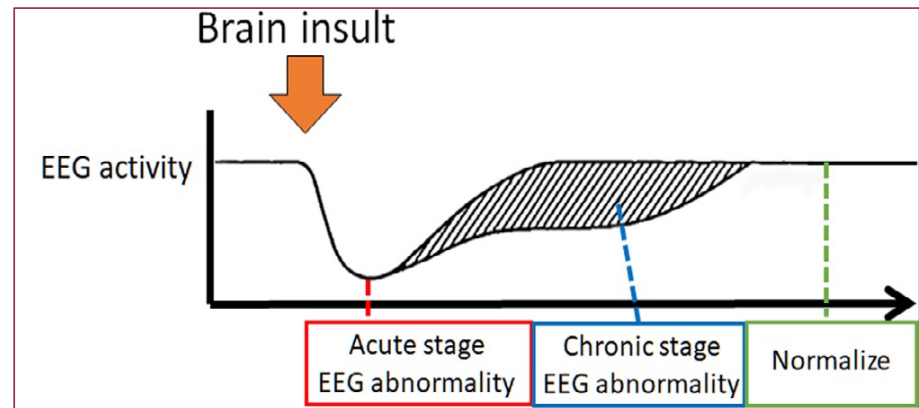


# *Indications for Neonatal EEG*

- To determine whether or not reported clinical events are seizures.
- To confirm diagnosis of aEEG events suspected to be seizures.
- To classify seizure type +/- aid in the diagnosis of specific syndromes.
- For monitoring neonates at risk for seizures in the absence of clinically evident seizures.
  - neonates with HIE with/without therapeutic hypothermia
  - vascular events including ischemic stroke, intracranial haemorrhage
  - preterm <32w CGA with additional risk factors
  - clinical encephalopathy (other than HIE)
  - post operative newborn heart surgery for congenital heart disease and babies on ECMO
  - intracranial infection
  - inborn errors of metabolism
- To assess for seizure control after treatment and to confirm resolution of seizures.
- For the assessment of interictal background patterns as part of risk stratification for evolving brain injury and prediction of acute seizures and for neurological prognosis

# Neonatal EEG for prognosticating outcome

- EEG may indicate prognosis for neurodevelopmental outcome after
  - HIE and other hypoxic brain injury
  - Acute symptomatic seizures
  - Preterm brain injury
- Ideal timing in HIE day 2-3
  - Good outcome if normal < 8 hr
  - Poor outcome if
    - No sleep wake cycling >48 hr
    - Severe background abnormality >24 hr ( @48hrs when cooled)
    - Mild background abnormality > 3 weeks
  - No predictive value if normal or mildly abnormal day 5-21
- In preterm brain injury serial EEG has best predictive value at 30, 32 & 35weeks



## RECORDING STANDARDS

***Standard 1*** - Before starting testing the patient is identified, and the clinical information from the referral verified.

*Guideline 1.1* – Appropriate (video) consent should be taken if data is to be used for teaching or training.



## RECORDING STANDARDS

***Standard 2*** – Disposable cup electrodes should be applied, dependant on scalp access, with electrode impedances  $<10\text{k}\Omega$  and where possible, balanced across the scalp. Electrode impedance should be checked and recorded at the beginning and end of the recording (Lloyd *et al*, 2014; Tsuchida *et al*, 2013, Mafilâtre *et al*, 2020).

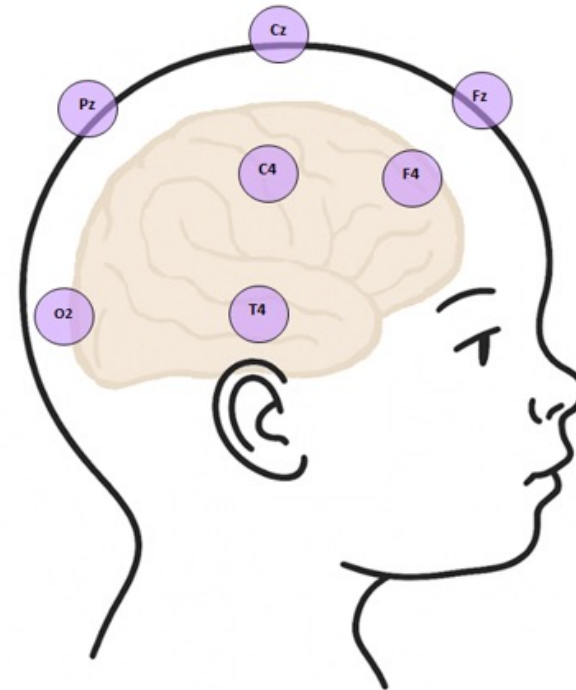
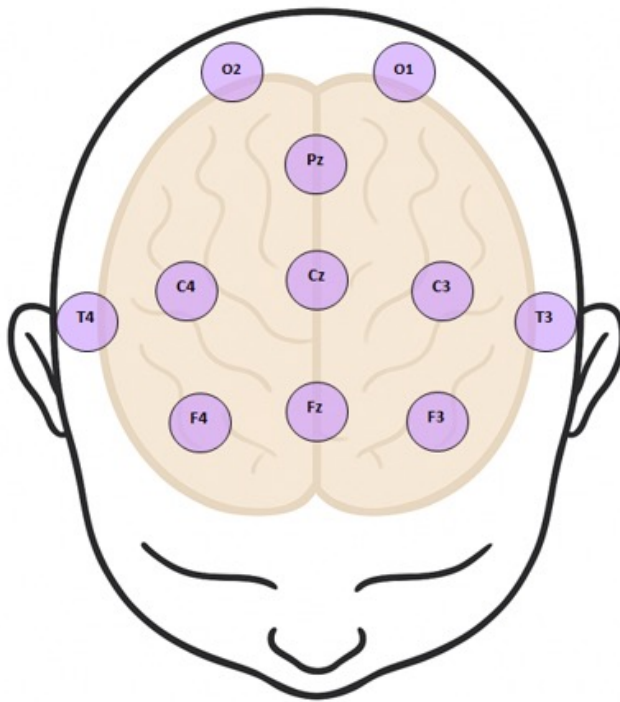


*Guideline 2.1* – A minimum of 13 (11 active; ground and ref) electrodes is recommended for patients  $>35\text{w}$  CGA on the NICU/PICU. This can be increased to a full 10-20 montage

*Guideline 2.2* - A minimum of 11 (9 active; ground and reference) electrodes is recommended where scalp access is limited due to prematurity ( $<35\text{w}$  CGA) or other factors.

*Option 2.1* – Support from the clinical team providing direct care (e.g. bedside nurse, neonatologist) can be enlisted to move the patient carefully during electrode application.

## Appendix: Suggested neonatal montage





***Standard 3 – Polygraphy should be included for the duration of the recording (Tsuchida et al, 2013; Malfilâtre et al 2021).***

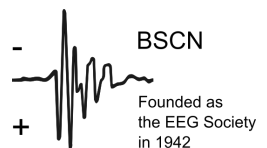
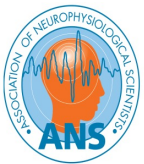
***As a minimum this should include:***

- **Bilateral deltoid EMG**
- **ECG**
- **Respiratory monitoring**

*Guideline 3.1 – Additional EMG channels should be considered, with positioning tailored to the clinical history (Tsuchida et al, 2013, André et al, 2010).*

*Option 3.1 – Consider integration of vital signs monitors.*

*Option 3.2 – Consider oxygen saturation monitoring (Tsuchida et al, 2013, Malfilâtre et al, 2020).*



		Electrode modality					
		EEG	ECG	EMG	Respiratory	EOG	SaO <sub>2</sub>
Settings	Electrode type	Disposable surface	Disposable surface	Disposable surface	Band or transducer	Disposable surface	Transducers
	LFF (Hz)	0.5	1.6	5	0.15	0.5	Parallel DC recording channel
	HFF (Hz)	70	30	300	15	30	
	Sensitivity (uV/cm)	Adjustable	Adjustable	Adjustable	Adjustable	Adjustable	

### **Standard 4 – Skin integrity is vulnerable in this age-group, particularly in pre-term infants**

(ASET Position Statement on Skin Safety During EEG Procedures – A Guideline to Improving Outcome, 2016; [El Ters et al, 2018](#); Ness, Davis, & Carey, 2013)

- Skin must be prepared using a gentle cleansing gel.
- Electrodes should be attached as per standard 2 using water-soluble adhesives and not using collodion adhesive.
- Electrodes should be removed using water; any skin irritation or sores should be highlighted to the bedside nursing team.



## RECORDING STANDARDS

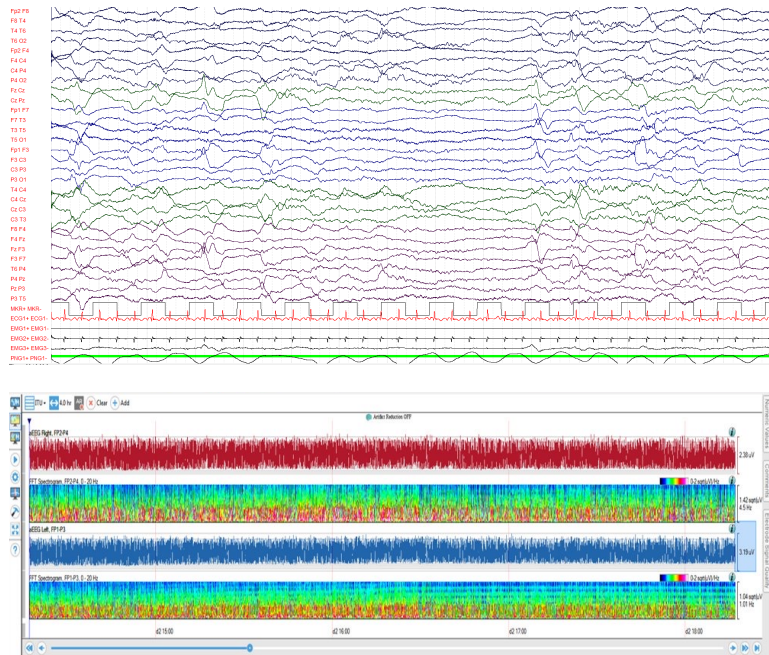
**Standard 5 - Recording duration should be a minimum of 60 mins, incorporating wake and sleep states (Shellhaas et al, 2011; Malfilâtre et al, 2020).**

*Guideline 5.1* – Consider prolonging recordings for event capture for clinically suspected seizures. For long recordings over 24 hr, observation of the scalp integrity should be noted, recorded & neonatal staff notified if damage is identified

*Option 5.1* - If the EEG is inactive/isoelectric, recording can be limited to 30 mins, ensuring adequate reactivity testing is performed in this time.

*Option 5.2* - Consider timing, increasing duration and repeat recordings for assessment of electrographic seizures and neurological prognosis according to the indication.

*Option 5.3* – Quantitative EEG may be used as an **adjunct** to assess seizure burden, effects of medication and subtle changes in background state, particularly in the muscle relaxed patient.



### **Standard 6 – Time-locked video should be recorded with full visibility of the patient (Shellhaas *et al*, 2011; Malfilâtre *et al*, 2020)**

*Guidelines 6.1* – Remove blankets for a clear view throughout the recording.

*Option 6.1* - A marker button or diary sheet may be used to allow accurate marking of clinical events (Shellhaas *et al*, 2011). Bedside staff and parents/carers should be encouraged to identify and describe events of concern.

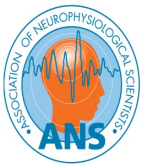
*Option 6.2* – Where vital signs information cannot be integrated into the recording, incorporating vital signs monitors into the video capture frame may be useful.



### Standard 7 – Tactile stimulation

- In neonates of >32w CGA, tactile stimulation should be performed to assess reactivity unless this is contraindicated.
- Stimulation should be repeated, where possible, to confirm reproducibility and this should be accurately documented in the recording (André *et al*, 2010). Repeat stimulation should be performed at an interval of 30 seconds OR when the EEG baseline returns if reactivity is observed, whichever is longer.
- A graded approach should be used Auditory -> tactile-> +/- suction (Hwang J et al 2022, ACNS and European guideline)

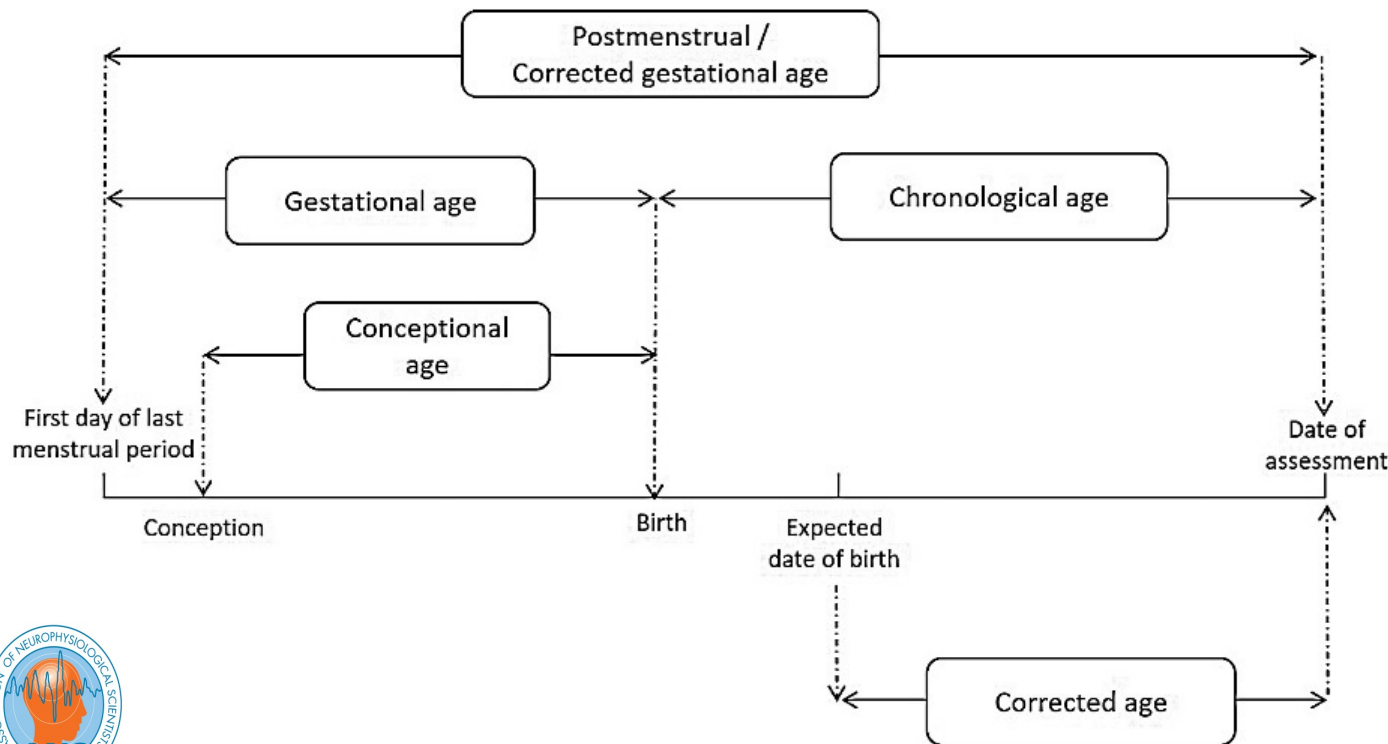
*Guideline 7.1* – Units/ departments should agree standardised local protocols to assess reactivity in conjunction with the neonatal team to ensure adherence to developmental care (see appendices for example stimulation protocol).



## REPORTING STANDARDS

### *Standard 1 – A concise and relevant clinical history should be documented, including:*

- CGA/PMA, GA



Born at GA 34+2 weeks



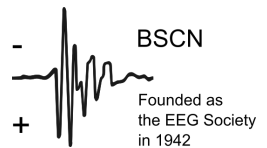
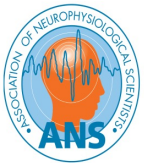
At 4 weeks of age (chronologic age)  
CGA or PMA 38+2 weeks



### ***Standard 1 – A concise and relevant clinical history should be documented, including:***

- CGA/PMA, GA
- Medication (anti-seizure medications, sedation (loading or maintenance))
- Indication for test
- History: Pregnancy, birth complications, APGAR score, cord pH, blood gas results
- Family history including genetic abnormalities and consanguinity
- Cooling – core temperature at time of EEG and timeframe to rewarming
- Event description including event frequency and time of last event
- Imaging results and previous EEG/aEEG/CFAM results if from

*Guideline 1.1 – Consider including results of other investigations.*



### *Standard 2 – A factual report should include:*

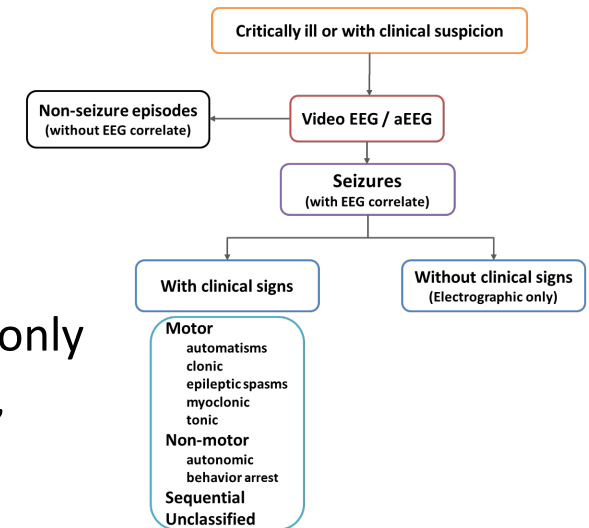
- Electrode array including polygraphy
- Technical factors including artefacts or changes in recording duration
- Presence/absence of sleep-wake cycling
- Background EEG description including:
  - Continuity – duration of continuity/discontinuity
  - Synchrony/symmetry in each patient state
  - Normal/abnormal graphical elements
- Description of events captured; habitual/non-habitual and associated EEG changes
- Description of stimulation and reproducibility
- Any medical interventions performed during the recording
- Autonomic changes recorded



## REPORTING STANDARDS

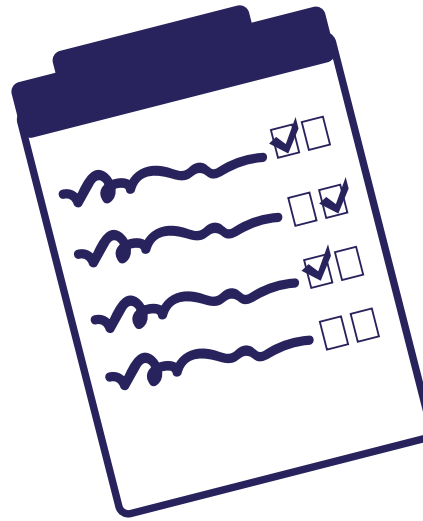
### *Standard 3 – A clinical interpretation should include:*

- A statement of whether the background is appropriate for GA, CGA / PMA
- Wake and sleep
- Effect of anaesthesia/ cooling etc
- A statement on normality or abnormality in the recording
- Presence/ absence of background abnormalities incl. location
- A summary of captured clinical, electroclinical and electrographic-only events with EEG correlates including duration, frequency, location, evolution and propagation
- Classification of electro-clinical seizures (Pressler et al, 2021)
- Comment on the interpretation and significance of the observed findings in the context of the clinical presentation. This may include reference to and/or recommendation for adjunct investigations
- A comparison with previous EEG records should be made where these are available



### *Standard 4 – Signing off*

- The professional status of the practitioners performing the investigation and providing the report are stated
- The report is signed by the practitioner taking medico-legal responsibility for it.



### **Standard 5 – Timing of reporting:**

- A written or verbal report should be disseminated to the requesting clinical team in a timely manner.
- For in-patients, this should be on the same day as test requested (Shellhaas *et al*, 2011).



### **Standard 6 – Storage of data :**

- Clinical and electrographic data should be stored in accordance with local guidelines.

# Thank you to Neonatal audit team

- Bryony Carr
- Daniela P. Quayle
- Emma Dean
- Gareth Payne
- Kelly Bill
- Khazina Waraich
- Matthew Sparkes
- Rachel Thornton
- Ronit Pressler
- Sushma Goyal
- Tatyana Yermakova

(In alphabetical order)

